

ENZYMATIC SYNTHESIS OF AZA-L-TRYPTOPHANS: The Preparation of 5- and 6-Aza-L-tryptophan¹

Milton J. Sloan and Robert S. Phillips*,
*Departments of Chemistry and Biochemistry,
University of Georgia, Athens, Georgia 30602.*

(Received 22 June 1992)

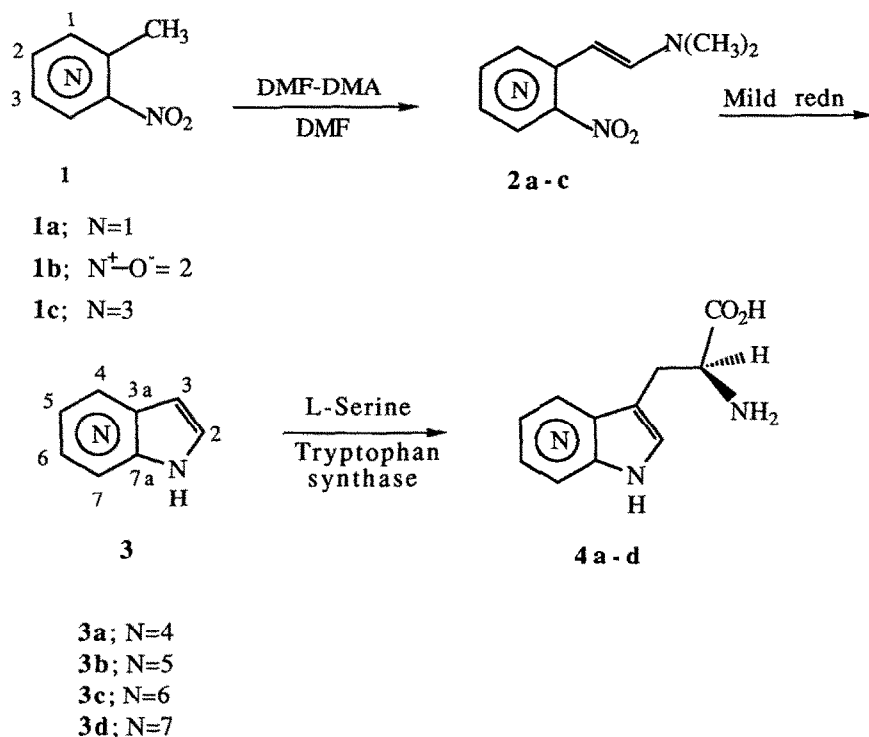
Abstract:

Enzymatic reaction of the azaindoles **3a-d** with tryptophan synthase [EC. 4.2.1.20] and L-serine results in synthesis of aza analogs of tryptophan **4a-d**. The synthesis of **4b** and **4c** has not been previously reported. The azaindoles **3a-c** were prepared from nitropicolines **1a-c**.

The essential amino acid tryptophan is vital to the normal function of all living organisms. It serves a central role in the biosynthesis and metabolism of proteins, peptides, enzymes, hormones, alkaloids, and other biologically important products. 7-Azatryptophan has shown a wide range of biological activities in a number of systems²⁻⁶. 7-Azatryptophan is a competitive antagonist of tryptophan metabolism, and has been used to study enzyme mechanisms⁷. It has been synthesized both chemically⁸ and enzymatically^{9,10} and is commercially available in racemic form. However, there have been few studies of other aza analogs of tryptophan^{6,9,11-13}. Consequently, the potential of other aza analogs of tryptophan to possess significant biological activity is of considerable interest. We report here a convenient synthesis of these aza analogs, and describe the first reports of 5- and 6-aza-L-tryptophan. Our method also provides for the preparation of 4-aza-L-tryptophan and improved yields of 7-aza-L-tryptophan⁹. Thus, we have prepared 4-, 5-, 6-, and 7-aza-L-tryptophan in one step from the corresponding azaindole with L-serine and recombinant tryptophan synthase¹⁴. The absence of C-2 and C-3 proton coupling in the NMR spectrum in each of the isomers prepared clearly indicates condensation on carbon at C-3, rather than on nitrogen as observed with indazole¹³.

7-Azaindole (Aldrich) was obtained commercially, and 6, 5, and 4-azaindole were prepared by modifying the methods reported by Yakhontov et al. ^{15,16}, and by Dormoy and Heymes¹⁷. Condensation of *ortho*-nitropicolines **1a-c** with N,N-dimethylformamide dimethylacetal (DMF-DMA) gave the enamines **2a-c** which upon reduction gave **3a-c**.

SCHEME 1



The reported catalytic hydrogenation and Raney nickel-hydrazine reduction of the enamines **2a-c** did not give satisfactory results in our hands. These methods afforded fair amounts of uncyclized byproducts in which the olefin bond was reduced prior to the nitro group. A milder and more selective method using catalytic hydrogen transfer was found to give much higher yields and cleaner products¹⁸. Five percent palladium on carbon in a formic acid-methanol mixture or aqueous ammonium formate was used to effect cyclization to the azaindoles **3a-c**. Aqueous ammonium formate was found to be more suitable than formic acid-methanol, since the workup was facilitated by sublimation of the ammonium carbonate byproduct during the course of the reaction. The azaindoles were then added to a reaction mixture of L-serine and tryptophan synthase¹⁹. Isolated yields of 61-88% were obtained after 2-11 days²⁰. We are presently investigating the reactivity of each azaindole in the reaction catalysed by tryptophan synthase.

Acknowledgements:

We wish to acknowledge support by the National Institutes of Health (grant No. GM42588-03 to RSP) and to the University of Georgia Graduate School (to MJS). The culture of *E. coli* CB149 containing pSTB7 was kindly provided by Dr. Edith W. Miles.

REFERENCES AND NOTES

1. Reported in part at the 42nd Southeast/46th Southwest Combined Regional Meeting of the American Chemical Society: New Orleans, LA, December, 1990; Abstract No. 429; and the 43rd Southeast Regional Meeting of the American Chemical Society, Richmond, VA; November, 1991; Abstract No. 253.
2. G. W. Kidder, & V. C. Dewey, *Biochem. Biophys. Acta.* **1955**, *17*, 88.
3. A. B. Pardee, & L. S. Prestidge, *Biochem. Biophys. Acta.* **1958**, *27*, 330.
4. M. R. Robinson, & B. L. Robinson, *J. Am. Chem. Soc.* **1955**, *77*, 456.
5. A. B. Pardee, V. G. Shora & L. S. Prestidge, *Biochem. Biophys. Acta.* **1956**, *21*, 406.
6. L. N. Yakhontov & A. A. Prokopov, *Uspekhi Khimii(Eng. Transl.)* **1980**, *49(5)*, 428.
7. R. S. Phillips, *J. Am. Chem. Soc.* **1989**, *111*, 727.
8. V. A. Azimov M. Ya. Uritskaya & L. N. Yakhontov, *Khimiko-Farmatsevticheskii Zhurnal (Eng. Transl.)* **1968**, *2(11)*, 605.
9. M. Wilcox, *Anal. Biochem.* **1974**, *59*, 436.
10. *Chem. Abstr.* **1974**, *80*, 131,685b.
11. P. Gmeiner & J. Sommer, *Ach. Pharm.(Weinheim)* **1988**, *321*, 505.
12. H. R. Snyder, C. B. Thompson & R. L. Hinman, *J. Am. Chem. Soc.* **1952**, *74*, 2009.
13. H. Tanaka, K. Tanizawa, T. Arai, K. Saito, T. Arai & K. Soda, *FEBS Lett.* **1986**, *196(2)*, 357.
14. The enzyme was obtained from cell extracts produced from a multicopy plasmid, pSTB7, containing the *trpA* and *trpB* genes from *Salmonella typhimurium*, and expressed in *Escherichia coli* CB149. (E. W. Miles, H. Kawasaki, S. A. Ahmed, H. Morita, H. Morita, & S. Nagata, *J. Biol. Chem.* **1989**, *264*, 6280).
15. A. A. Prokopov & L. N. Yakhontov, *Khim. Geterotsikl. Soedin. (Eng. Transl.)* **1977**, 1135.
16. V. A. Azimov & L. N. Yakhontov, *Khim. Geterotsikl. Soedin. (Eng. Transl.)* **1977**, 1145.
17. J. R. Dormoy & A. Heymes, **1985**, French Patent FR2,564,836Cl. C07D471/04; *Chem. Abstr.* **1986**, *105*, 115044w.
18. a) B. El Amin, G. M. Anantharamaiah, G. P. Royer & G. E. Means, *J. Org. Chem.* **1979**, *44*, 3444; b) H. Wiener, J. Blum & Y. Sasson, *J. Org. Chem.* **1991**, *56*, 4481; c) S. Rajeswar, K. Drost & M. P. Cava, *Heterocycles*, **1989**, *29(3)*, 415; d) I. D. Entwistle, A. E. Jackson, R. A. W. Johnstone & Y. Sasson, *J. Chem. Soc. Perkin Trans. 1* **1977**, 443.
19. A typical reaction mixture consisted of azaindole (16.9 mM), L-serine (22.8 mM), pyridoxal phosphate (53 mM), potassium phosphate buffer (100 mM, pH 8), NaCl (16.2 mM), tryptophan synthase (2000-4000 units), and NaN₃ (3 mM). The reaction mixture was incubated in the dark at 37°C. Progress was monitored by TLC, and the reaction was stopped by heating in a water bath at 80°C for approx. 30 min. to denature the protein. The precipitated protein was

removed by filtration or centrifugation, and the solution concentrated *in vacuo*. Isolation of **4a-c** was effected by applying the solution over reverse phase silica gel, eluting with 15% aqueous methanol (150 mL).

20. **4-Azatryptophan 4a** Isolated yield of 53 mg (61%) from 50 mg of 4-azaindole, after 2-3 days; $[\alpha]^{25}_{\text{D}} +10.7^{\circ}$ (c 0.5, 1N HCl); IR (nujol) 3307 cm^{-1} (br), 3137, 1684, 1622, 1590, 1558, 1506, 1404, 1333, 1306, 1234, 1158, 1133, 1114, 973, 889, 849, 761; ^1H NMR ($\text{D}_2\text{O}/\text{DCl}$, 300.13 MHz, acetone δ 2.06 ppm) 8.41 ppm(d, 1H, $J=8.6\text{Hz}$, 7-ArH), 8.35(d, 1H, $J=5.9\text{Hz}$, 5-ArH), 7.92(s, 1H, 2-ArH), 7.53(dd, 1H, $J=5.9\text{Hz}$, $J=8.4\text{Hz}$, 6-ArH), 4.35(t, 1H, $J=6.5\text{Hz}$, αCH), 3.50(dd, 1H, $J=15.9\text{Hz}$, $J=6.5\text{Hz}$, βCH), 3.42(dd, 1H, $J=15.9\text{Hz}$, $J=6.5\text{Hz}$, βCH); ^{13}C NMR ($\text{D}_2\text{O}/\text{DCl}$, 75.46 MHz, acetone δ 30.6 ppm) 171.75 ppm($\text{C}=\text{O}$) 136.36(5-C) 135.03(3a-C or 7a-C) 134.21(6-C) 133.18(7a-C or 3a-C) 129.19(7-C) 117.22(2-C) 103.06(3-C) 53.03(αCH) 24.43(βCH_2); Anal for $\text{C}_{10}\text{H}_{11}\text{N}_3\text{O}_2\cdot\text{H}_2\text{O}$ Calcd %C 53.81, H 5.87, N 18.82; Found %C 54.26, H 5.51, N 18.74. **5-Azatryptophan 4b** Isolated yield of 202 mg (77.5%) from 150 mg of 5-azaindole after 10-11 days; $[\alpha]^{25}_{\text{D}} +21.6^{\circ}$ (c 0.5, 1N HCl); IR (nujol) 3122 cm^{-1} (br), 1609(s), 1576, 1541(s), 1540, 1400, 1337, 1318, 1236, 1207, 1166, 1091, 1028(s), 881, 822, 811, 656.3; ^1H NMR ($\text{D}_2\text{O}/\text{DCl}$, 250.13 MHz, D_2O 4.65 δ ppm) 8.86 ppm(s, 1H, 4-ArH), 8.10(d, 1H, $J=6.7\text{Hz}$, 6-ArH), 7.72(d, 1H, $J=6.7\text{Hz}$, 7-ArH), 7.56(s, 1H, 2-ArH), 3.99(t, 1H, $J=6.0\text{Hz}$, αCH), 3.34(d, 2H, $J=6.0\text{Hz}$, βCH); ^{13}C NMR ($\text{D}_2\text{O}/\text{DCl}$, 62.89 MHz,) 173.86 ppm($\text{C}=\text{O}$) 134.78(4-C) 131.97(6-C) 130.90(7-C) 130.15(7a-C or 3a-C) 124.21(3a-C or 7a-C) 111.61(3-C) 109.83(2-C) 54.88(αCH) 26.22(βCH_2); Anal for $\text{C}_{10}\text{H}_{11}\text{N}_3\text{O}_2\cdot 2.5\text{H}_2\text{O}$ Calcd %C 47.99, H 6.59, N 16.79; Found %C 47.35, H 6.52, N 16.04. **6-Azatryptophan 4c** Isolated yield of 337 mg (88%) from 200 mg of 6-azaindole after 6-7 days; $[\alpha]^{25}_{\text{D}} +25.0^{\circ}$ (c 0.615, 1N HCl); IR (nujol) 3385 cm^{-1} (br), 3121, 1620, 1567, 1421, 1400, 1377(s), 1363, 1346, 1314, 1282, 1246, 1136, 1113, 1090, 1067, 1044(s), 966(s), 917(s), 899, 835, 769; ^1H NMR ($\text{D}_2\text{O}/\text{DCl}$, 300.13 MHz, acetone δ 2.06 ppm) 8.80 ppm(s, 1H, 7-ArH), 8.02 (d, 1H, $J=6.6\text{Hz}$, 5-ArH), 7.92(d, 1H, $J=6.6\text{Hz}$, 4-ArH), 7.93(d, 1H, $J=1.0\text{Hz}$, 2ArH), 4.24 (t, 1H, $J=6.1\text{Hz}$, αCH), 3.41(d, 2H, $J=6.1\text{Hz}$, βCH); ^{13}C NMR ($\text{D}_2\text{O}/\text{DCl}$, 75.46 MHz) 172.81 ppm($\text{C}=\text{O}$) 139.35(7-C) 137.73(5-C) 132.32(7a-C or 3a-C) 128.77(4-C) 128.02(3a-C or 7a-C) 116.40(2-C) 110.53(3-C) 54.47(αCH) 25.99(βCH_2); Anal for $\text{C}_{10}\text{H}_{11}\text{N}_3\text{O}_2\cdot 2\text{H}_2\text{O}$ Calcd %C 49.78, H 6.22, N 17.42; Found %C 49.78, H 6.20, N 17.39. **7-Azatryptophan 4d** Isolated yield of 30.6 mg (88%) from 20 mg of 7-azaindole after 3-4 days; $[\alpha]^{25}_{\text{D}} +14.7^{\circ}$ (c 0.275, 1N HCl); ^1H NMR ($\text{D}_2\text{O}/\text{DCl}$, 300.13 MHz, acetone δ 2.06 ppm) 8.43 ppm(dd, 1H, $J=8.0\text{Hz}$, $J=1.0\text{Hz}$, 6-ArH), 8.13(dd, 1H, $J=6.0\text{Hz}$, $J=1.0\text{Hz}$, 4-ArH), 7.41(s, 1H, 2-ArH), 7.33(dd, 1H, $J=8.0\text{Hz}$, $J=6.0\text{Hz}$, 5-ArH), 3.95(t, 1H, $J=6.20\text{Hz}$, αCH), 3.27(d, 2H, $J=6.20\text{Hz}$, βCH_2); ^{13}C NMR ($\text{D}_2\text{O}/\text{DCl}$, 75.46 MHz, acetone δ 30.6 ppm) 173.87 ppm($\text{C}=\text{O}$) 139.27(7a-C or 3a-C) 136.53(6-C) 133.76(5-C) 128.58(4-C) 125.47(3a-C or 7a-C) 115.77(2-C) 109.76(3-C) 54.74(αCH) 26.03(βCH_2).